Getting started with Qsep₁-Plus

Document No.: F0080 Ver.: CHT&ENG-B

Sample Trays

(8, 12 and 16+3 wells)



Packing List $Qair_{box}$ & Power Adapter Qsep₁-Plus C105200 (S1 & S2 Cartridge Kit) Qsep₁-Plus Power Adapter

Ethernet Cable

C105200 (S1 & S2 Cartridge Kit)

SD Card & Software Key

Install Qsep₁-Plus

1. Unpack $\mathit{Qair}_{\mathit{box}}$ and tighten the air tube with connector

2. Plug power cord into $Qair_{box}$

(* Please check the $\mathit{Qair}_\mathit{box}$ label on the power cord)

Mini Centrifuge



3. Remove the protecting foam before turning on Qsep₁-Plus





* Scan the QR code for instruction video



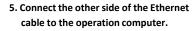
4. Plug the power cord, Ethernet cable and the other side of the air tube into Qsep₁-Plus and turn on the $Qair_{bax}$ (Please make sure the label on the power cord is $Qsep_i$)







Your *Qsep*₁-Plus is READY TO USE





*DO NOT switch on the instrument immediately after powering off Wait at least 5 seconds

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- 1. Ensure the SD card is properly inserted in the socket (Fig. 1 and 2).
- *SD card needs to be inserted before powering on.
 The SD card CANNOT be initialized with hot plugging.



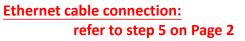


- 2. Connect the air tube.
- 3. Plug the power cord (labeled Qsep₁-Plus)
- Switch on Qsep₁-Plus, the LED lights up (multiple color light) (Fig. 3). Wait for 25 seconds until the instrument and Wi-Fi initialized, and the green LED will start flashing (Fig. 4).
- *DO NOT switch on the instrument immediately after powering off. Wait at least 5 seconds





*The green flashing LED shows the instrument is connected. If not, please repeat the step 4.





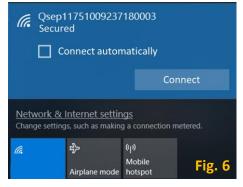
<u>Wi-Fi connection</u>: follow the steps below to connect with *Qsep*₁-Plus



Check if the AP source is available from your Wi-Fi network in the computer (Fig. 5).

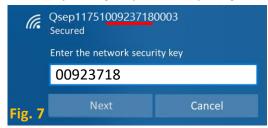


Find the SSID named with "Qsep1+instrument ID (16 digits)" and click "Connect" (Fig. 6)



Enter the password:

The password will show in the sticker which is behind the instrument. It is the same as the middle 8 digits of instrument serial number (same as SSID name) by blinding the prefixal "Qsep1" (Fig. 7).



*If the computer cannot find the SSID of $Qsep_1$ -Plus, keep the distance between $Qsep_1$ -Plus and the computer within 10 meters and start from step 4 again.

- *If the computer cannot connect with Qsep_-Plus
- Please check the wi-fi IP assignment is "Automatic (DHCP)"
- Allow Q-Analyzer pass through firewall or turn off the firewall



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- 5. Check the LED status.
- 6. Double click the icon of *Q-Analyzer*.





7. Q-Analyzer user interface:



- 7-1. Click "Search", and the instrument ID will display.

 *Please confirm the instrument ID is the
 connected with instrument
- 7-2. Click "Connect", the *Qsep*₁-Plus image will change the color when it's connected.

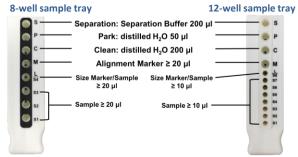


Packing List of Cartridge Kit (Cat. C105200)



Cartridges
Alignment Maker
Separation Buffer
Dilution Buffer
Mineral Oil
Buffer Tray
Dropper
0.2 ml tubes

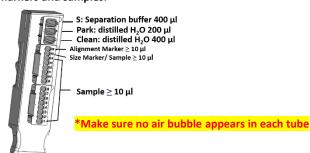
8. For 8-well and 12-well sample tray, prepare buffer and Alignment Marker in 0.2ml PCR tubes and allocate at the corresponding position on the sample tray.



*For 12-well sample tray, user needs to use 0.1ml tube (C104252) for sample and size marker *S/P/C well can only place 0.2ml PCR tubes



For 16+3-well sample tray, please use 16+3 Sample Tube (C104254) and follow the instruction below to load the buffers, markers and samples.



- Click "Open Sample Door", and the sample door will open automatically.
- 10. Slide the buffer tray into the instrument. Make sure sample tray is pushed to the end.



- 11. Click "Park", and the sample tray will move into instrument.
- 12. Open the cartridge door by pressing the white button and insert cartridge (guiding groove facing front).
 - *Please follow cartridge unpacking guide to unpack the cartridge before using





13. Close the cartridge door and click "Latch".



*The cartridge information will be displayed after latch.





Operation Quick Start

14. Cartridge Calibration:

New cartridge need to be calibrated before use. Please follow the steps below to proceed.



14-1. Click "HV Check".



* The storage and transportation condition may influence the Gelmatrix and cause unstable current. During HV Check, check current (gray line) and see if it's stable. if current is unstable, please repeat this step 2-3 times.





During the "HV Check", the last LED will show red light

Blue → SD card

Green → WiFi connectable
Red → HV check

Green Red

14-2. Click "Calibrate".



*The "Calibrate" button will only display after HV check



Please check the Alignment Marker has been placed in correct position.

Software will recognize peaks from Alignment Marker signal. DO NOT use Size Marker or DNA sample doing "Calibrate".

* For troubleshooting, please refer to cartridge unpacking guide for details.

15. Click on the blank column and designate ① the sample locations, ② test method, sample duration, runs, ③ result name and ④ Para by following steps 15-1, 15-2, 15-3 and 15-4.

		Sequenc	e	pen	Sav	10	Save As		
SN	Sample Position	Method	Sample Duration	Runs	Separation Duration		Result Name	Para	[Add]
1				1		None		_	Delete
	1	2					3	4	Up
									Down
									00:00
П	Micro Vial								

15-1. Click the "Sample Position", mark the position of sample on the plate and then click "OK".

s a O	Separation Buffer	Edit Clear All Save FI
p 🗓 👵	Park	Sample ID Sample Decorption Sample Name Sample Date Patient Name Patient
9	Clean/Wash Buffer	84
M 💿	Alignment Marker 20-1K	53 SALE(#1)
S4 (Size Marker C109200 -	sarrela 2 S2
S3 🧶	Sample	Sarrole 1
S2	Sample	
S1 🧑	Sample	
	- Compre	
	_	

15-2. Click "Method" to select analysis method.

Method Selector					
Application	C DNA	c	RNA	C Protein	
Analysis Type	@ Qualitativ	C Quantitative Sample V	olume(x):	и	
Alignment Marker	[₹ 20-1K(S	316) 🔻 20	1000 ← Reduce №	Normal C Enhance	
Cartridge Type	S2 ▼ Sta	ndard Cartridge(Shelf Life: 6 N	onths)		
Sample Concentration	High (Frag	pment >10 ng/ul) (2	Regular (Fragment: 0,1 ~ 10 ng/	ul) C Low (Fragment <0.1)	ng/ull)
Method		Description	Range	Remar	k
M-4-10-08-160		Sample Injection 4kv 10s Separation 8kv 160s	10bp~5000bp Best Resolution: 4bp~10bp	i .	
gDNA(NGS)		Sample Injection 4liv 10s Separation 8liv 200s		Sheared Genomic DI	LA:
T-HVPurge-08-120	0	Sel Refill with HV on for 120s			
T-Purge-120	(Sel Refill without HV for 120s			
1 "	1/1				
G High Voltage Pure	e C Pirne	Purge Modification			
					_
Customized Metho	4				0

* If you want to analyze the samples with Alignment Marker, please choose the appropriate Alignment Marker and then place it in the corresponding position. (Check ✓ the box)

Contact Information:

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15-3. Click on "Result Name" and enter the name of result files.

		Sequen	се	Open	Sa	ve Save As		
SN	Sample Position	Method	Sample		Separation Duration	Result Name		Para
		M-4-10-08-160				NGS Sample ID	¥	. /

15-4. Click the icon "Para" and set the parameters.
(Baseline Factor, Peak Threshold, Calculate, etc.).

Baseline Fac		
Calculate	Reference Marker Table	Browse
	Create Size Marker C109200(S15) ▼ □ Every 4 ▼ times	
	Size marker Injection time: Auto w sec(s)	
	Reference Marker Table:	
		Browse
Smear	C Distribution 100 16 C Range app	
Peak Calling		Browse
Auto Assign 1	88 288	
- Event Based	Show Report Setting	

16. Click "Run" to start the process.

	Qsep1: 7229009419530003
	Disconnect
	Open Sample Door
Dry, No.	Park

